

# Reporting and Evaluation of Studies of Biomarkers and Omics-based Predictors: *REMARK Guidelines and NCI Omics Checklist*

*Canadian Statistical Sciences  
Institute (CANSSI) Workshop*

Lisa McShane, PhD

Biometric Research Branch, DCTD

U.S. National Cancer Institute

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# Outline

- ❑ Background & definitions for tumor marker prognostic studies
- ❑ Role of reporting guidelines: REMARK
- ❑ Scaling up to omics-based predictors: cautions in study design and conduct
- ❑ Criteria to judge readiness of omics-based test to be used in a clinical trial
- ❑ Summary remarks

# Definitions

## □ Biomarker

<http://www.cancer.gov/dictionary>: “Biological molecule found in blood, other body fluids, or tissues that is a sign of a normal or abnormal process, or of a condition or disease.”

## □ Prognostic

Associated with clinical outcome in absence of therapy (natural course) *or with standard therapy all patients are likely to receive*

- May or may not be relevant for therapy decisions

**FOCUS: Tumor prognostic markers**

# State of the Tumor Marker Literature

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JOURNAL OF CLINICAL ONCOLOGY

ASCO SPECIAL ARTICLE

## American Society of Clinical Oncology 2007 Update of Recommendations for the Use of Tumor Markers in Breast Cancer

*Lyndsay Harris, Herbert Fritsche, Robert Menzel, Larry Norton, Peter Ravdin, Sheila Taube, Mark R. Somerfield, Daniel F. Hayes, and Robert C. Bast Jr*

Purpose: To update the recommendations for the use of tumor marker tests in the prevention, screening, treatment, and surveillance of breast cancer.

“ . . . primary literature is characterized by studies that included **small patient numbers**, that are **retrospective**, and that commonly perform **multiple analyses until one reveals a statistically significant result**. . . many tumor marker studies **fail to include** **how patients were treated or analyses of the marker in different treatment subgroups**. The Update Committee hopes that adherence to . . . **REMARK criteria** will provide more informative data sets in the future.

# State of the Tumor Marker Literature

“Studies of ‘prognostic’ markers of no real future clinical utility and single biomarker studies will not be considered. Reports of studies into prognostic markers should be prospective and have a clear view of the practical clinical applications of the results. Retrospective analysis of biomarkers can be considered, if done within the framework of data collected from a prospective trial, with appropriate statistics and with multivariate analysis that includes established predictive/prognostic markers. Reports of prognostic tumor marker studies should follow the REMARK guidelines (available from [www.equator-network.org](http://www.equator-network.org)).”

J. B. Vermorken

Editor-in-Chief

Statement of editorial intent

*Annals of Oncology* 2012; 23:1931-1932

# REMARK: REporting guidelines for tumor MARKer prognostic studies

Lisa M. McShane, Douglas G. Altman, Willi Sauerbrei, Sheila E. Taube, Massimo Gion, and Gary M. Clark for the Statistics Subcommittee of the NCI-EORTC Working Group on Cancer Diagnostics (*J Natl Cancer Inst* 2005; 97:1180-1184, and simultaneously in *BJC, EJC, JCO, NCPO*)

## Recommended reporting elements to facilitate

- ❑ Evaluation of **appropriateness & quality** of study design, methods, and analysis
- ❑ Understanding of **context** in which conclusions apply
- ❑ **Reproducibility**
- ❑ **Comparisons** across studies, including formal meta-analyses

# REMARK: Target Studies

- ❑ Studies relating marker values to clinical events (e.g., recurrence, death, response)
- ❑ NOT primarily aimed at biological discovery studies, but use encouraged to extent possible
  - Patients
  - Specimens
  - Assays
- ❑ NOT sufficient for studies developing multiplex classifiers/risk scores (e.g., derived from omics data), but applicable to studies assessing them

# REMARK Elements: Introduction

- ❑ State all marker(s) examined
- ❑ Study objectives
- ❑ Pre-specified hypotheses

# Common Tumor Marker Study Design



What can we do with  
our marker on these  
89 specimens?

- ❑ “Convenience” specimens
- ❑ Heterogeneous patient characteristics
- ❑ Treatments: Unknown, non-randomized, not standardized
- ❑ Insufficient sample size (underpowered)
- ❑ Uncertain specimen and data quality

# REMARK Elements: Materials & Methods

## □ Patients

- Inclusion/exclusion (e.g., stage, subtype), source, treatments

## □ Specimen characteristics

- Format, collection, preservation, storage
- See BRISQ criteria (Moore et al, *Cancer Cytopathology* 2011; 119:92-101)

# REMARK Elements: Materials & Methods (cont.)

## □ Assay methods

- Detailed protocol (reagents/kits), quantitation, scoring & reporting, reproducibility, blinding

Example: Systematic review (43 studies) of Ki67 in early breast cancer (Stuart-Harris et al, *The Breast* 2008; 17:323-334)

- English publication, Jan. 1995 – Sept. 2004
- $\geq 100$  patients, OS or DFS endpoint
- Results
  - 7 different antibodies for IHC, single or combination
  - 19 different cutpoints, ranging from 0-30%
  - Significant between-study heterogeneity and evidence for publication bias

# REMARK Elements: Materials & Methods (cont.)

- Study design
  - Case selection (e.g., random, case-control), clinical endpoints, variables considered, sample size
- Statistical analysis methods
  - Models, variable selection, handling of missing data, multiple testing adjustments, validations

# Importance of identifying exploratory statistical analyses

EUROPEAN JOURNAL OF CANCER 43 (2007) 2559–2579



ELSEVIER

available at [www.sciencedirect.com](http://www.sciencedirect.com)



journal homepage: [www.ejconline.com](http://www.ejconline.com)



## Almost all articles on cancer prognostic markers report statistically significant results

Panayiotis A. Kyzas<sup>a</sup>, Despina Denaxa-Kyza<sup>a</sup>, John P.A. Ioannidis<sup>a,b,c,\*</sup>

<sup>a</sup>Clinical and Molecular Epidemiology Unit, Department of Hygiene and Epidemiology, University of Ioannina School of Medicine, Ioannina, Greece

<sup>b</sup>Biomedical Research Institute, Foundation for Research and Technology-Hellas, Ioannina, Greece

<sup>c</sup>Institute for Clinical Research and Health Policy Studies, Department of Medicine, Tufts-New England Medical Center, Boston, USA

“If you torture the data long enough they will confess to anything.”

*Source unknown*

# Statistical Analysis: Multiple Testing

- Multiple markers
- Multiple endpoints
- Multiple subgroups
- Multiple marker cutpoints
- Multiple models with multiple variables

Example: 8 subgroups defined by 3 binary factors

Number of independent tests ( $\alpha = 0.05$ per test)	Probability observe $\geq 1$ statistically significant ( $p < 0.05$ ) result
1	0.05
2	0.10
3	0.14
4	0.19
5	0.23
6	0.26
7	0.30
8	0.34
9	0.37
10	0.40

# Statistical Analysis: Cutpoint optimization

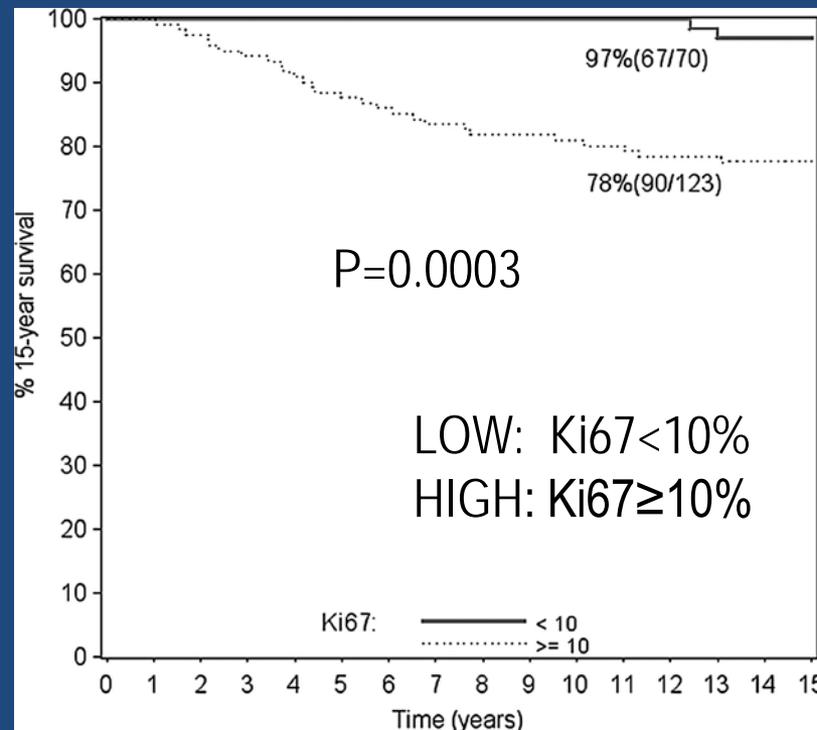
- ❑ 203 patients with lymph node negative primary breast cancer
- ❑ Proliferation marker Ki67 measured by IHC on 193/203
- ❑ No adjuvant systemic therapy (chemo or endocrine)

Endpoint = Breast  
Cancer Specific Survival  
(BCSS)

LOW: Ki67 < 10%  
15-yr BCSS = 97%

HIGH: Ki67 ≥ 10%  
15-yr BCSS = 78%

BCSS for low and high Ki67



# Statistical Analysis: Cutpoint optimization

Number of deaths, sensitivities and specificities according to a range of cut-off values of Ki67

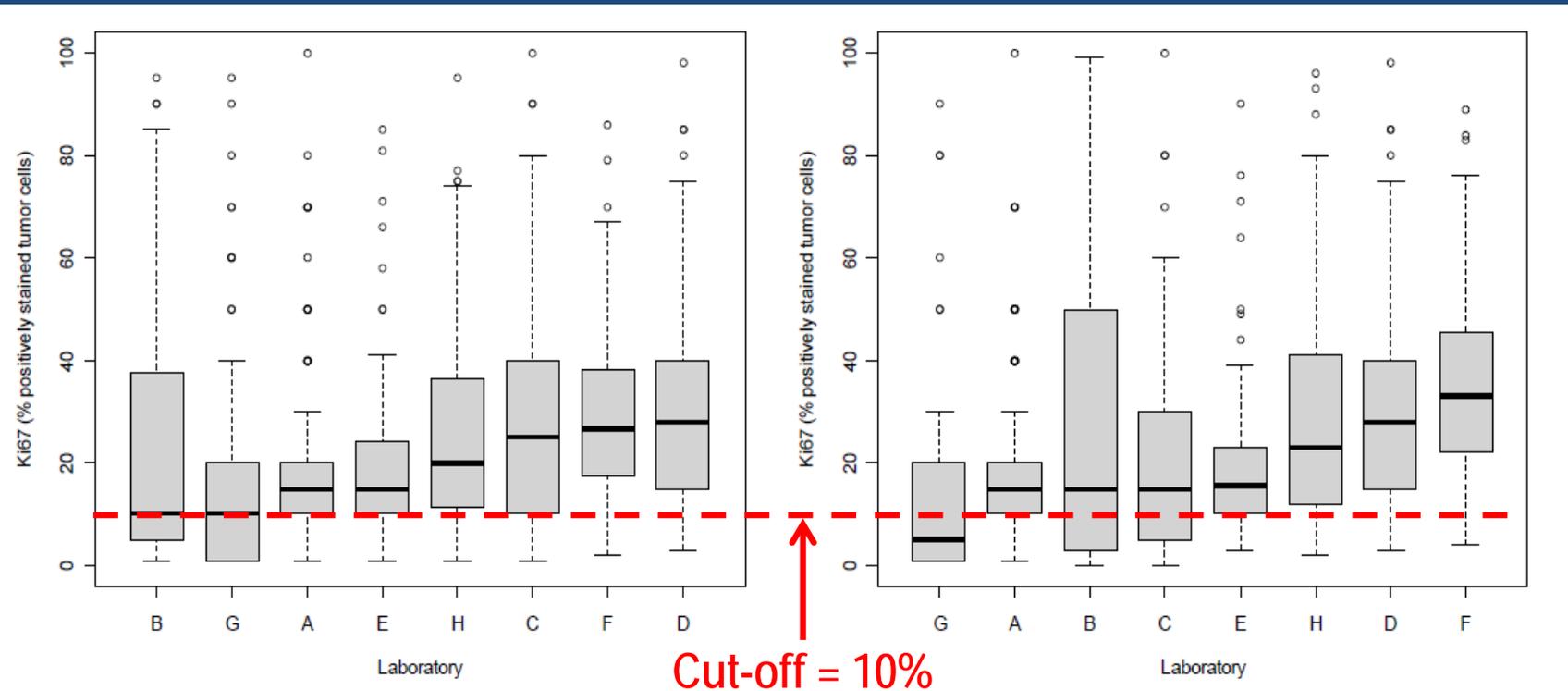
Ki67	No. died (%)	No. in category	Sensitivity	Specificity	Youden's index (J)
≥0	29 (15.0)	193	1	0	0
≥5	28 (17.6)	159	0.966	0.201	0.167
≥10	27 (22.0)	123	0.931	0.415	<b>0.346</b>
≥15	20 (21.3)	94	0.690	0.549	0.238
≥20	16 (22.5)	71	0.552	0.665	0.216
≥30	12 (25.0)	48	0.414	0.780	0.194
≥40	10 (27.8)	36	0.345	0.841	0.186
≥50	8 (27.6)	29	0.276	0.872	0.148

$$J = \text{Sensitivity} + \text{Specificity} - 1$$

Pathmanathan N et al. J Clin Pathol 2014;67:222-228 (Table 1)

# Statistical Analysis: Cutpoint optimization and impact on assay transportability

Side-by-side boxplots of Ki67 distributions with 8 labs assessing different TMA sections of same set of 100 breast cancer cases



Centrally stained, locally scored  
Median range: 10% to 28%

Locally stained, locally scored  
Median range: 5% to 33%

Polley M et al, J Natl Cancer Inst 2013; 105: 1897-1906 (Figure 2)

# REMARK Elements: Results

## □ Data

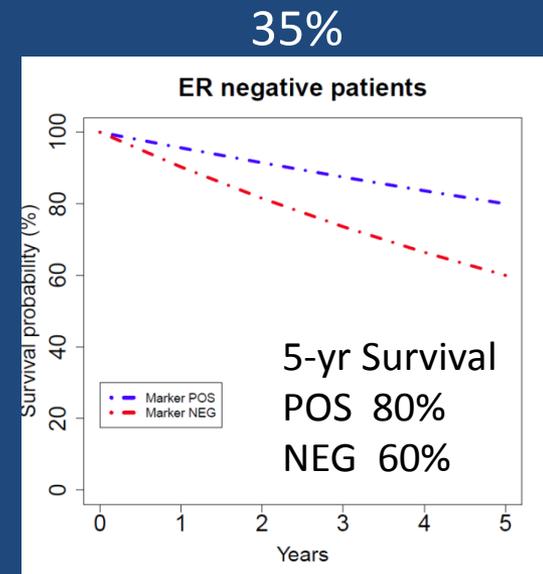
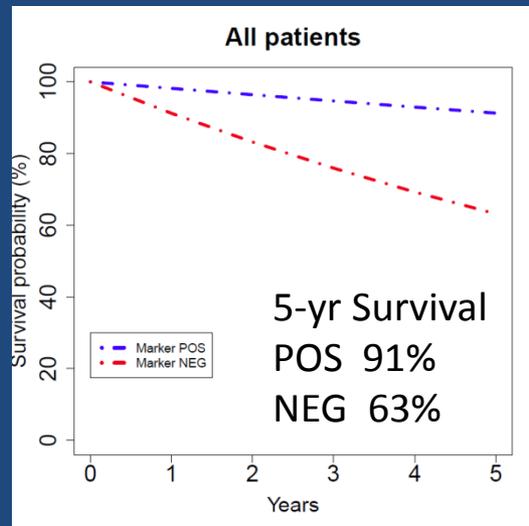
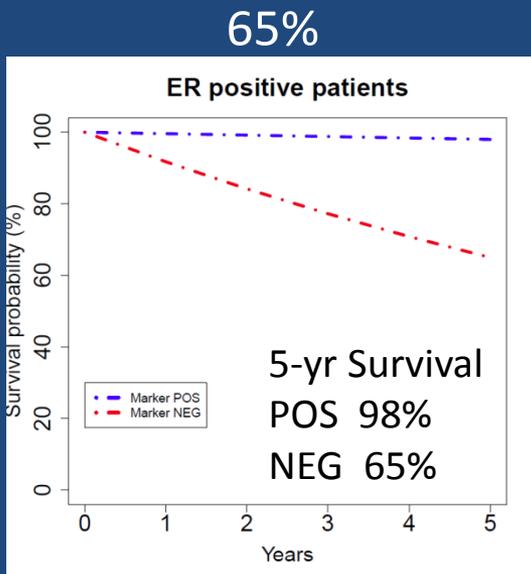
- Numbers of patients and events
- Demographic characteristics
- Standard prognostic variable distribution
- Tumor marker distribution

## □ Analysis & presentation

- Univariate analyses (marker vs. standard prognostic variables, marker vs. outcome)
- Multivariable analyses (association of marker with outcome after adjustment for standard prognostic variables)
- Measures of uncertainty for reported effect estimates

# REMARK Elements: Results (cont.)

- Multivariate analysis vs. subgroups
  - Subgroup analyses may be important for interpretation
  - Better yet, study more clinically homogenous populations



## REMARK Elements: Discussion

- ❑ Interpretation in context of pre-specified hypotheses
- ❑ Relevance to other studies
- ❑ Limitations
- ❑ Future research
- ❑ Clinical value

# REMARK Status & Future

- Explanation & Elaboration: Altman et al, *PLoS Medicine* 2012; 9(5):e1001216 (also *BMC Medicine* 2012; 10:51)
- “Before vs. after” reporting quality
  - Before: Mallett et al, *British Journal of Cancer* 2010; 102: 173-180
  - After: Underway
- Journals stating REMARK adherence requirements: *Ann Oncol, Breast Cancer Res Treat, Clin Cancer Res, J Clin Oncol, J Natl Cancer Inst, J Pathol*

# Scaling up to omics-based predictors

## □ Omics

“A term encompassing multiple molecular disciplines, which involve the characterization of global sets of biological molecules such as DNAs, RNAs, proteins, and metabolites.”

## □ Omics-based test

“An assay composed of or derived from multiple molecular measurements and interpreted by a fully specified computational model to produce a clinically actionable result.”

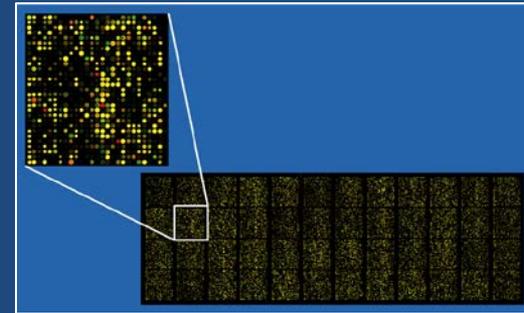
(Mathematical model component referred to as a predictor or classifier with outputs such as risk score or categorization.)

Institute of Medicine report: Evolution of Translational Omics  
<http://www.iom.edu/Reports/2012/Evolution-of-Translational-Omics.aspx>

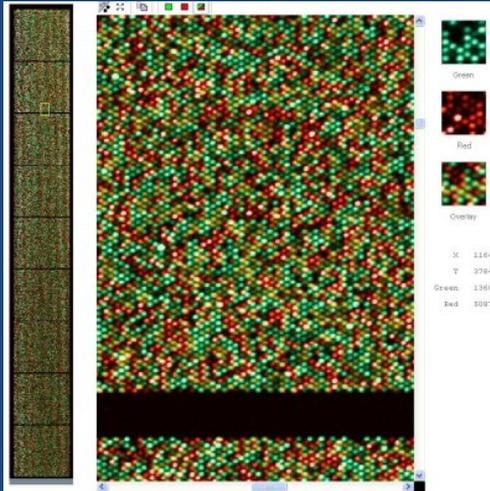
# Omic assays



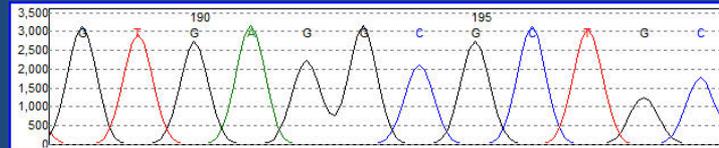
Affymetrix expression GeneChip



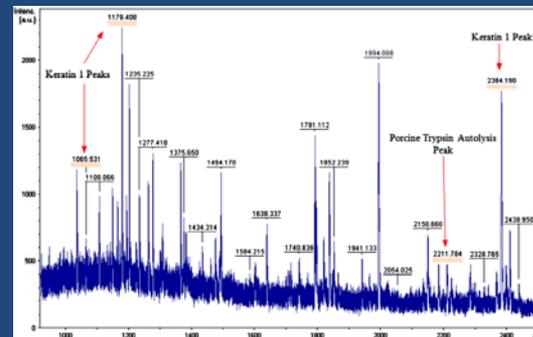
cDNA expression microarray



Illumina SNP bead array



Mutation sequence surveyor trace

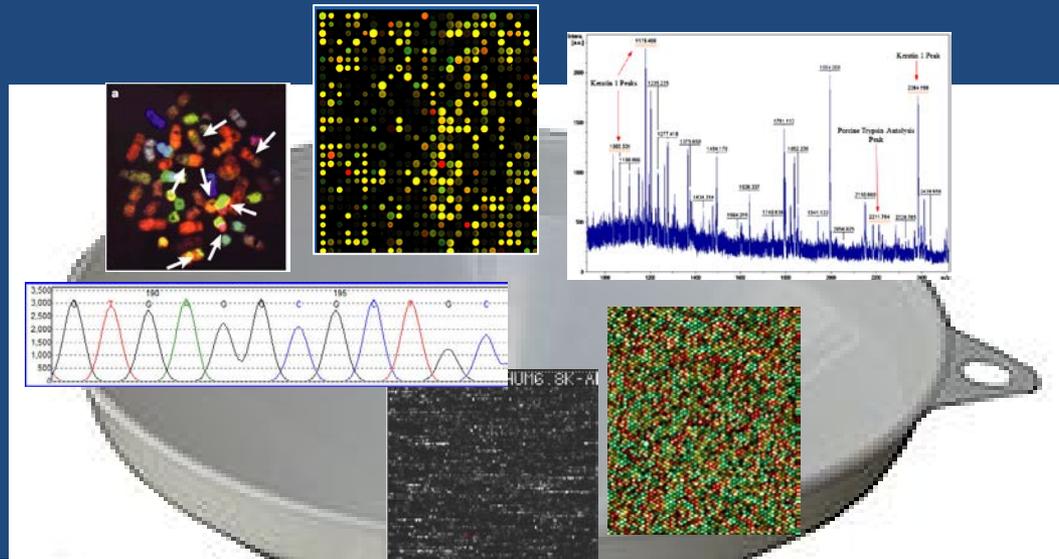


MALDI-TOF proteomic spectrum

# Translation from omics discoveries to clinically useful omics-based tests

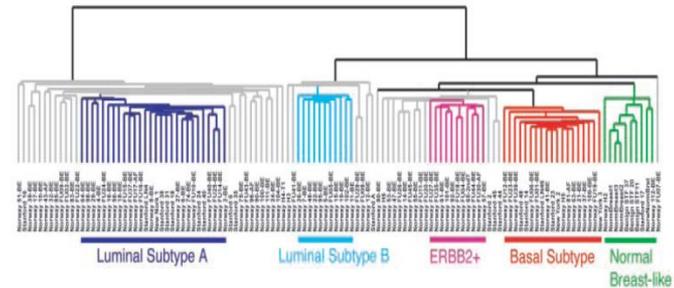
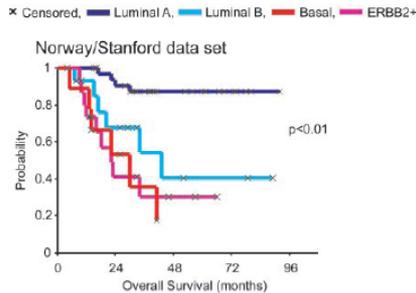
## High-throughput omics assays

Discovery



**Computational models**

**Predictors, classifiers, risk scores**



Clinical Utility?

# Paradigm for development of a clinically useful omics-based test

## Discovery

### Clinical validity

The test result shows an association with a clinical outcome of interest.

### Analytical validity

The test's performance is established to be accurate, reliable, and reproducible.

### Clinical utility

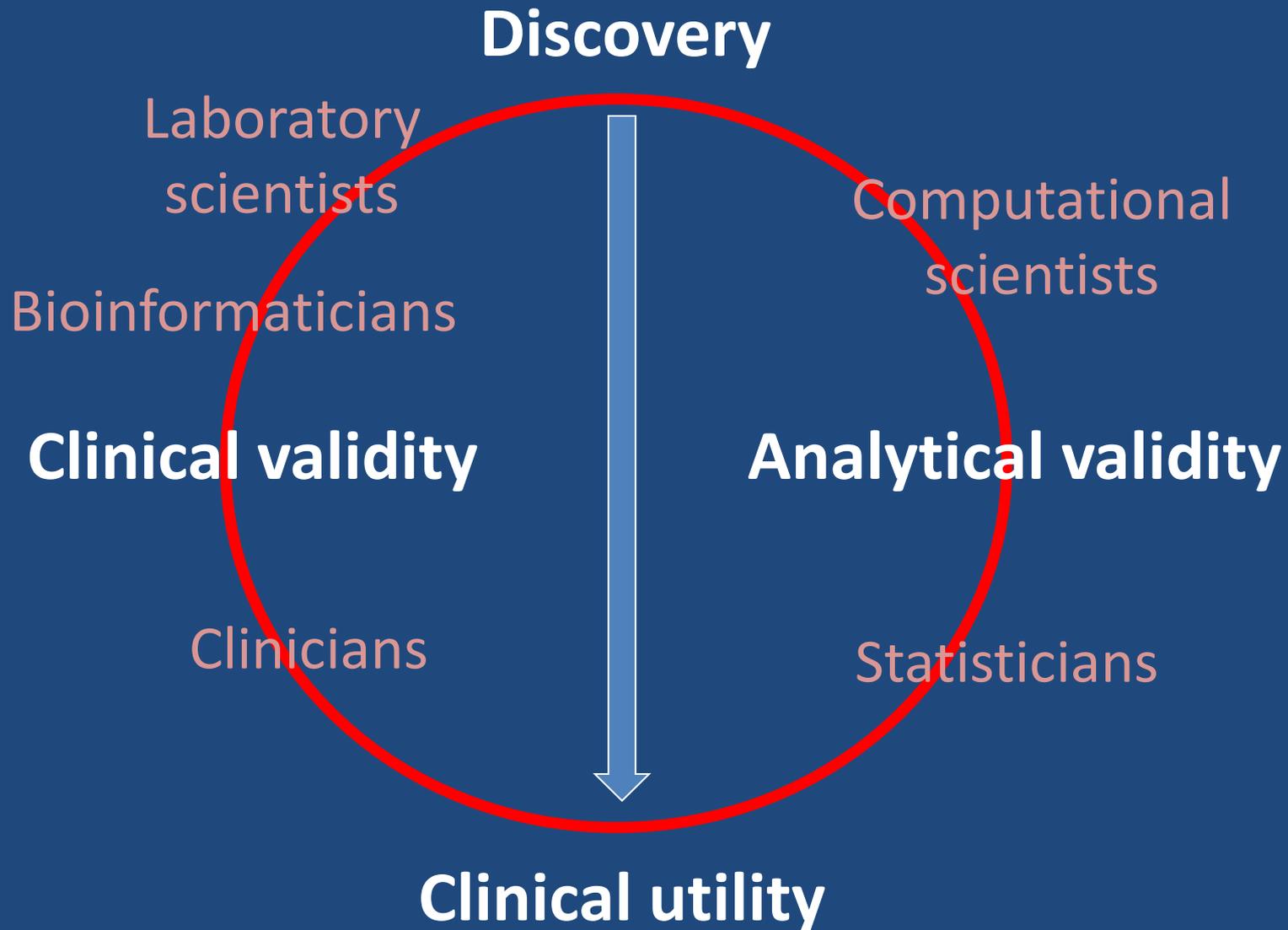
Use of the test results in a favorable benefit to risk ratio for the patient

Teutsch et al, *Genet Med* 2009;11:3-14

Simon et al, *J Natl Cancer Inst* 2009;101:1446-1452

McShane & Hayes, *J Clin Oncol* 2012;30:4223-4232

# It takes a collaborative team to go from discovery to clinically useful omics test



# NCI Criteria for the use of omics-based predictors in clinical trials

- Focus: Tests based on potentially complex mathematical models incorporating large numbers of measurements from omics assays
- Goals:
  - Make omics test development more efficient, reliable, and transparent
  - Avoid premature clinical implementation of omics-based tests

McShane et al, *Nature* 2013;502:317-320

McShane et al, *BMC Medicine* 2013;11:220

# Omics checklist divided into 5 domains

- ❑ Specimens
- ❑ Assays
- ❑ Model development, specification & preliminary performance evaluation
- ❑ Clinical trial design
- ❑ Ethical, legal, and regulatory

# Domain 1: Specimens

- ❑ Collection, processing & storage
- ❑ Specimen quality screening
- ❑ Minimum required amount
- ❑ Feasibility of collecting needed specimens
  - Achievable in standard clinical settings
  - Study/sample size planning

# Domain 1: Specimens example

- ❑ Statisticians can provide guidance in planning feasibility assessments and quality monitoring schemes to avoid disasters

## Example:

- Analysis of first 100 biological specimens collected in a large diagnostic study showed that only 20% were of adequate quality to be analyzable by the assay
- Problem traced to failure to promptly freeze the specimens after collection

## Domain 2: Assays

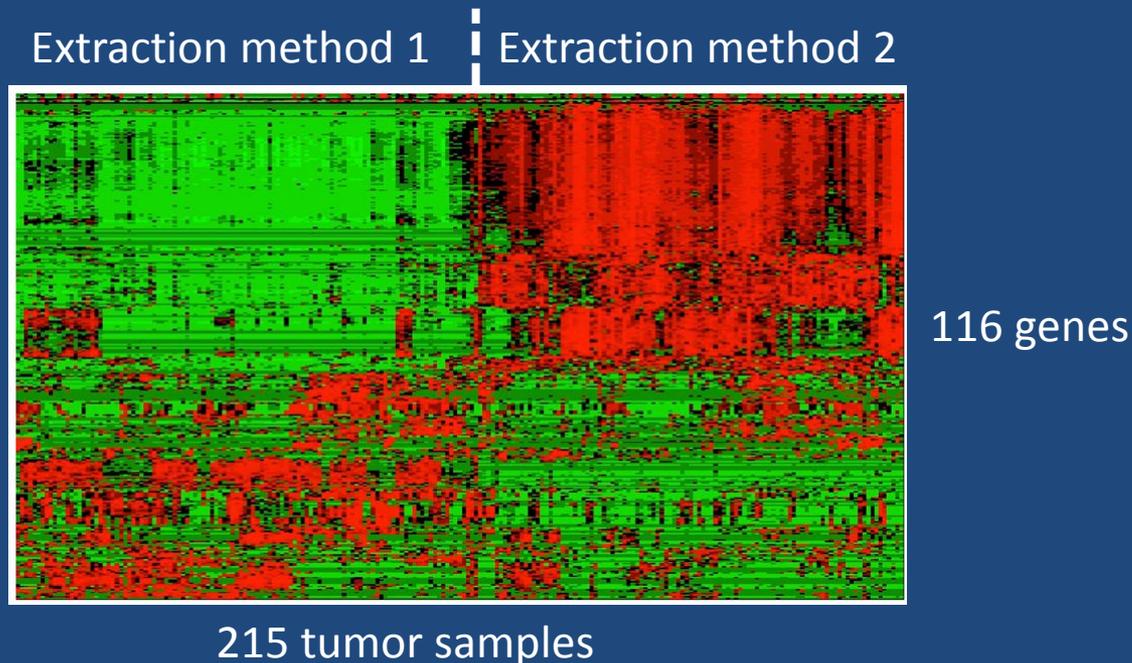
- ❑ Impact of changes in assay procedures
- ❑ Lock down SOP
- ❑ Quality criteria for assay values
  - Bad specimens, batch effects, equipment malfunction
- ❑ Analytical performance evaluation
  - Pennello, *Clinical Trials* 2013;10: 666–676
  - Jennings et al, *Arch Pathol Lab Med* 2009;133: 743–755
- ❑ Quality monitoring
- ❑ Turnaround time

# Domain 2: Assay example

- Assess impact of changes in any assay procedures, reagents, or equipment

## Example:

Dramatic effect of change in RNA extraction procedure on tumor gene expression microarray profiles, additional minor effect due to reagent changes by microarray manufacturer



# Domain 3: Model development & evaluation

- ❑ Quality of data (clinical & omics) used to develop and validate predictor models
- ❑ Appropriate statistical approaches for model development and performance assessment
- ❑ Intended use - data from clinically relevant patient population

# Domain 3: Data quality & batch effects

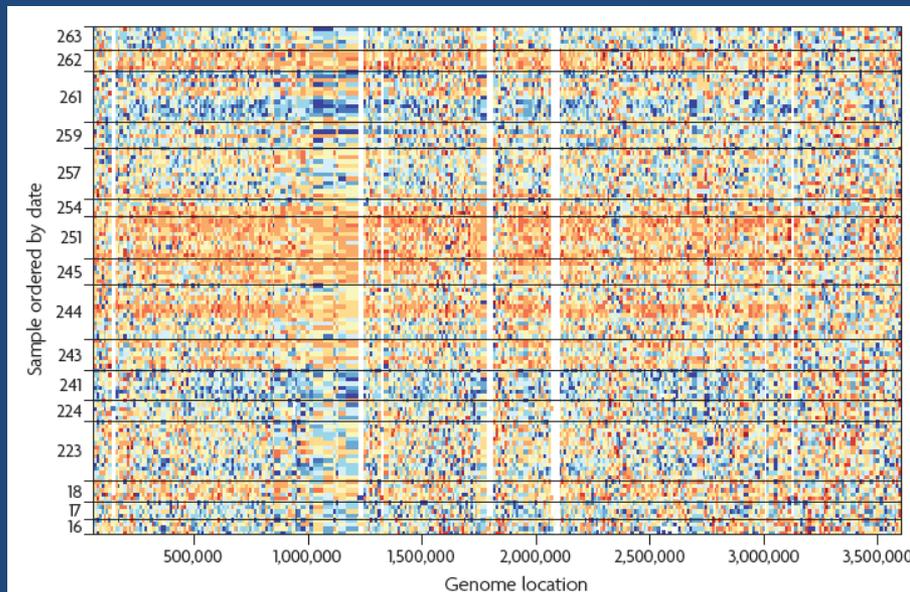
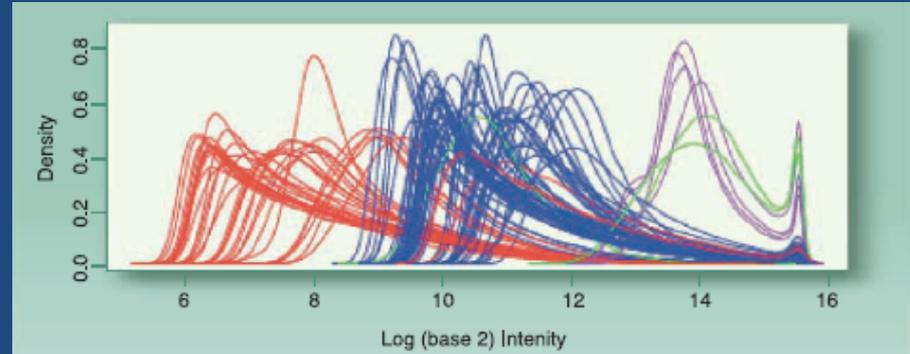
Density estimates of PM probe intensities (Affymetrix CEL files) for 96 NSCLC specimens

Red = batch 1

Blue = batch 2

Purple & Green = outliers?

(Owzar et al, *Clin Cancer Res* 2008;14:5959-5966)



Batch effects for 2<sup>nd</sup> generation sequence data (stand. coverage data).

Same facility & platform.

Horizontal lines divide by date.

(Leek et al, *Nature Rev Genet* 2010;11:733-739)

**BATCH EFFECTS ARE ESPECIALLY PROBLEMATIC IF CONFOUNDED WITH KEY EXPERIMENTAL FACTORS OR ENDPOINTS.**

# Domain 3: Dangers of overfitting

- A statistical model is **OVERFIT** when it describes random error (noise) instead of the true underlying relationship
  - Excessively complex (too many parameters or predictor variables )
  - Generally has poor predictive performance on an independent data set

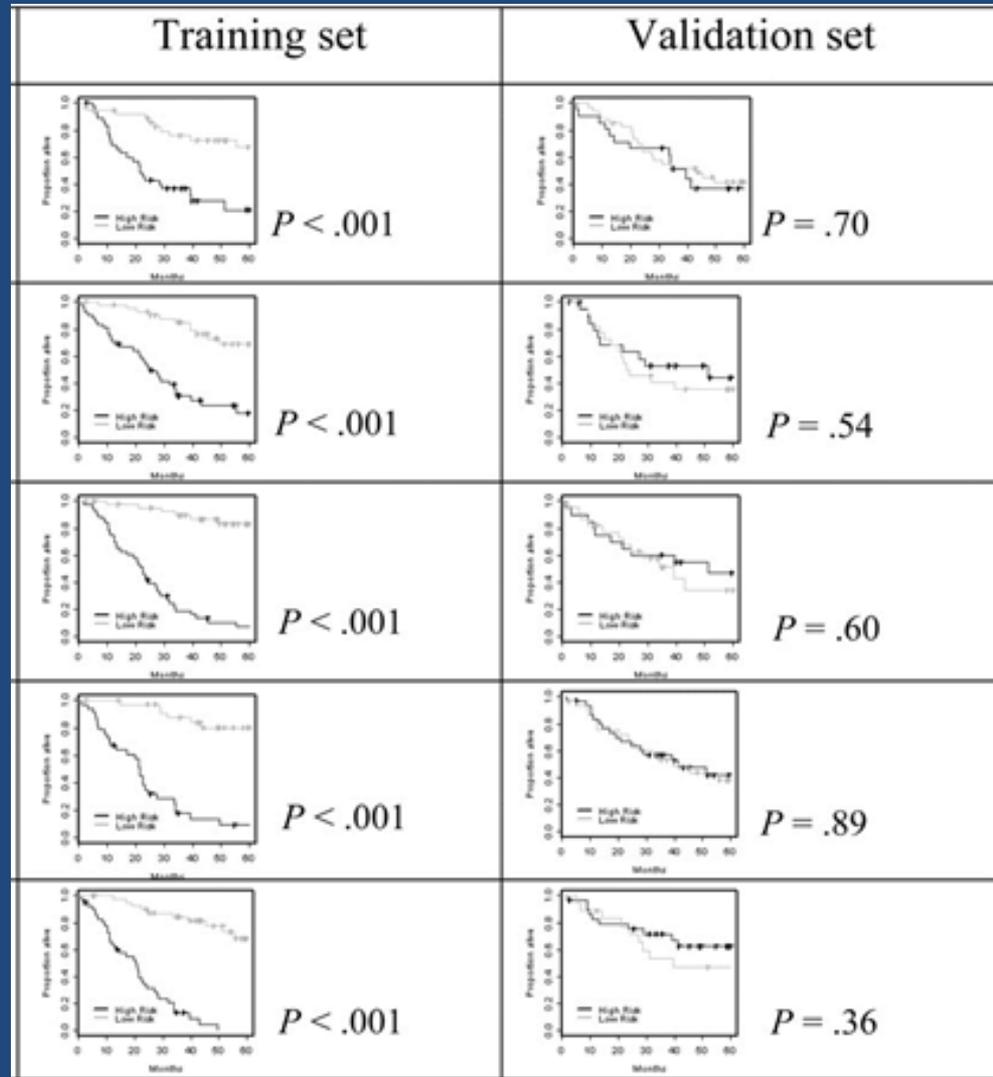
# Domain 3: Failure to detect overfitting

- **RESUBSTITUTION** is the naïve practice of evaluating performance of a model by “plugging in” exact same data used to build it
  - Seriously biased estimates of predictor performance
  - Overfitting will not be detected

# Domain 3: Avoid overfitting & resubstitution

## Simulation of bias in resubstitution estimates of predictor performance

- Goal: Develop prognostic signature from gene expression microarray data
- Survival data on 129 lung cancer patients (prior study)
- Expression values for 5000 genes generated randomly from  $N(0, I_{5000})$  (“noise”) for each patient
- Data divided randomly into training and validation sets
- Prognostic model developed from training set and used to classify patients in both training and validation sets (supervised principal components method)



(Subramanian & Simon, *J Natl Cancer Inst* 2010;102:464-474)

# Domain 3: Detection and avoidance of model overfitting

## □ Internal validation by use of data resampling techniques

- Split sample (training & test sets)
- Cross-validation
- Bootstrapping

Molinaro et al, *Bioinformatics* 2005;21:3301-3307

## □ External validation

- Assessment of predictor performance on a completely independent data set

## □ Model regularization techniques reduce, but don't completely eliminate overfitting

# Domain 3: Subtle forms of model overfitting

- ❑ Partial resubstitution
- ❑ Combining training and test sets
- ❑ Resubstitution with covariate adjustment
- ❑ Resubstitution comparison

Simon et al, *J Natl Cancer Inst* 2003;95:14-18

Subramanian & Simon, *J Natl Cancer Inst* 2010;102:464-474

Simon & Freidlin, [Correspondence] *J Natl Cancer Inst* 2012;103(5):445

Subramanian & Simon, *Contemporary Clinical Trials* 2013;36:636–641

McShane & Polley, *Clinical Trials* 2013;10:653-665

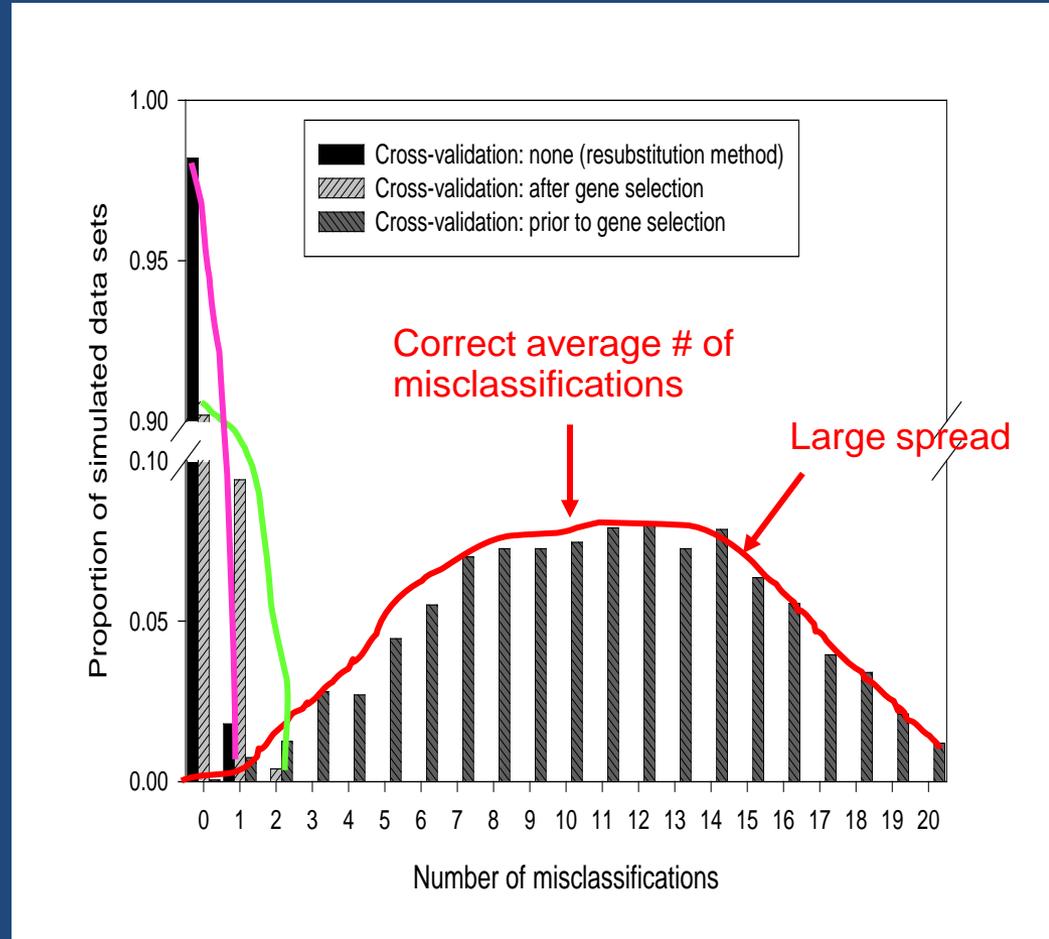
# Domain 3: Avoid partial resubstitution

Simulation experiment: 20 specimens; expression levels of 6000 genes randomly generated (Gaussian noise); arbitrary split of specimens into two groups of 10

## Prediction Method:

- Compound covariate
- Use 10 most differentially expressed genes to build classifier
- Calculate number of misclassifications

**Repeat simulation  
2,000 times**



Simon et al, *J Natl Cancer Inst* 2003;95:14-18

# Domain 3: Avoid combining training & test sets

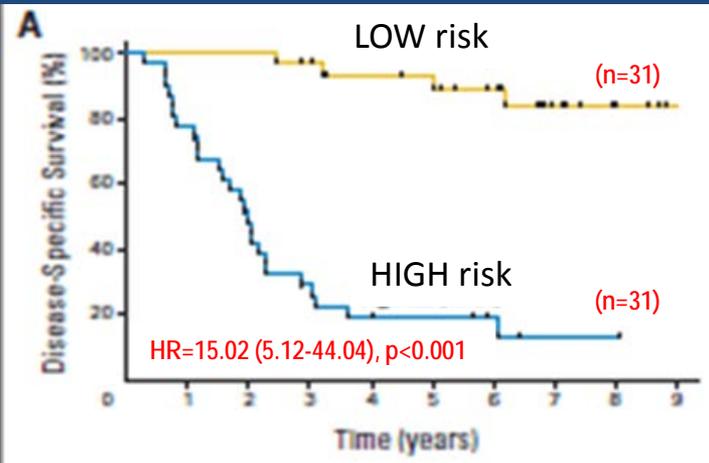
## Multivariable Model for Overall Survival (Training and Test sets combined)

Variable	HR	95% CI	P
Genomic score	2.43	1.94 – 3.06	< 0.001
Stand. molec. factor 1	1.77	1.41 – 2.22	< 0.001
Stand. molec. factor 2	0.66	0.48 – 0.93	0.02
Age group, $\geq 60$ yrs vs < 60 yrs	2.22	1.76 – 2.79	< 0.001

Combining Training data (used to develop genomic score) with Test data destroys the validation and interpretability of the adjusted effects

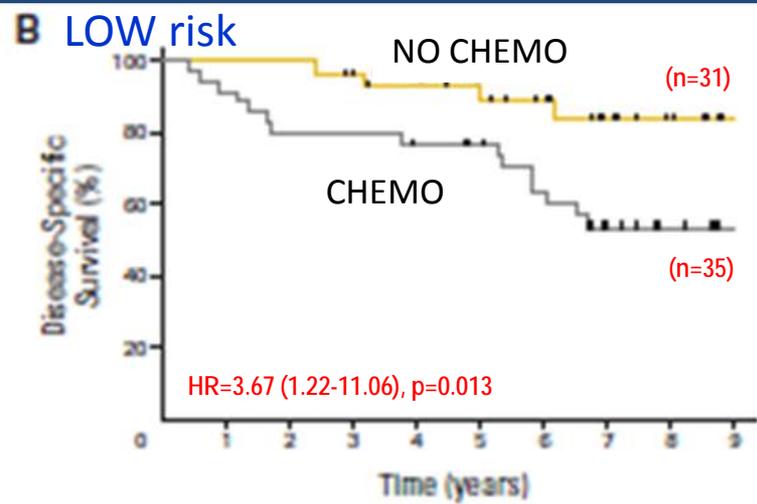
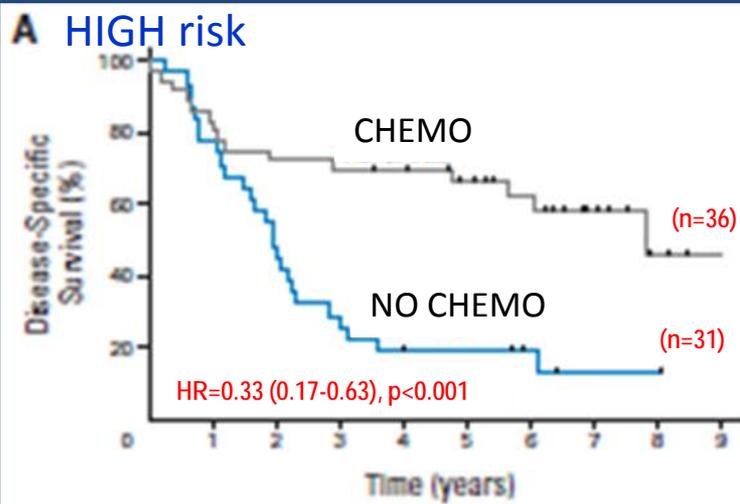
Nowhere in the paper was a multivariate analysis based solely on the Test set presented.

# Domain 3: Avoid comparisons with resubstitution estimates



Simon & Freidlin, [Correspondence]  
J Natl Cancer Inst 2012;103(5):445

Prognostic classifier fit using gene expression microarray data from clinical trial arm on which patients received no adjuvant chemotherapy (resubstitution)



Does the genomic predictor identify groups of patients who benefit differently from adjuvant chemotherapy?  
Can't conclude anything.

# Domain 3: Requirements for a rigorous validation of a predictor

- ❑ The predictor to be tested must be completely **LOCKED DOWN** and there must be a **PRE-SPECIFIED PERFORMANCE METRIC**. The lockdown includes all steps in the data pre-processing and prediction algorithm.
- ❑ The **INDEPENDENT VALIDATION DATA** should be generated from specimens collected at a different time, or in a different place, and according to the pre-specified collection protocol.
- ❑ Assays for the validation specimen set should be run at a different time or in a different laboratory but according to the **IDENTICAL ASSAY** protocol as was used for the training set.
- ❑ The individuals developing the predictor must remain completely **BLINDED** to the validation data.
- ❑ The validation **DATA SHOULD NOT BE CHANGED** based on the performance of the predictor.
- ❑ The **PREDICTOR SHOULD NOT BE ADJUSTED** after its performance has been observed on any part of the validation data. Otherwise, the validation is compromised and a new validation may be required.

# Domain 3: Fully-specified “locked down” predictor

Need all of the following:

- ❑ List of individual variables
- ❑ Data pre-processing steps (e.g., normalization/standardization of raw data)
- ❑ Equation/algorithm to make predictions
- ❑ Produces same or highly similar result when *same* data are input multiple times
- ❑ Predictor can be applied *one* case at a time

## Domain 3: Examples of predictors *not* locked down

- Example #1: List of variables (e.g., genes, proteins) with no indication of how to combine the variables
- Example #2: Data pre-processing using data from a collection of specimens (e.g., each gene expression value is standardized across a collection of cases as  $z = (x - \bar{x})/s$ )

How to pre-process data from a single new case?

Need to lock down pre-processing parameters or use reference set.

## Domain 3: Examples of predictors *not* locked down (cont.)

- Example #3: Use of ranks or percentiles
  - Linear combination scores computed on training set and classified using median score for the training set as cutpoint for classification of the training set cases
  - Linear combination scores computed on test set and classified using median score for the *test* set as cutpoint for classification of the *test* set

Cutpoint may shift from data set to set due to assay batch or cohort effects.

How is a single new case classified?

## Domain 3: Example of predictors *not* locked down (cont.)

- Example #4: “Black-box” computer programs that produce varying predictions when run multiple times on same data
  - Stochastic model averaging methods
  - Methods that employ clustering methods with random initial centroids (e.g., some implementations of K-means clustering)

Example: Same data from  $\approx 100$  cases input twice, 20% chance of flipping (low/high risk) prediction, run to run

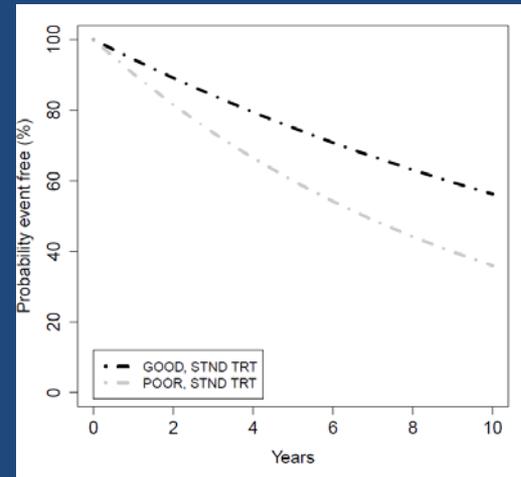
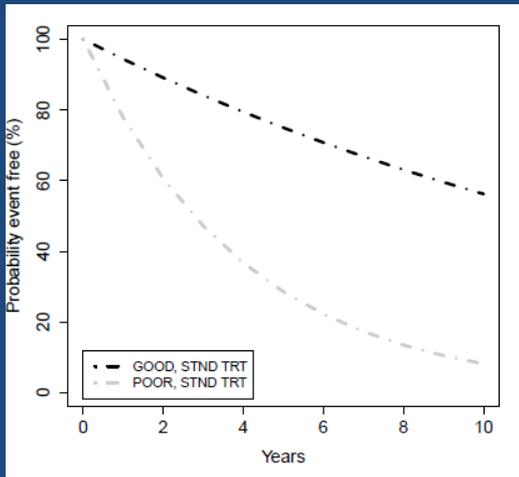
Either varying aspects must be locked (e.g., fix random number seed), or it must be established that variation across repeat runs is minimal.

# Domain 4: Clinical trial design

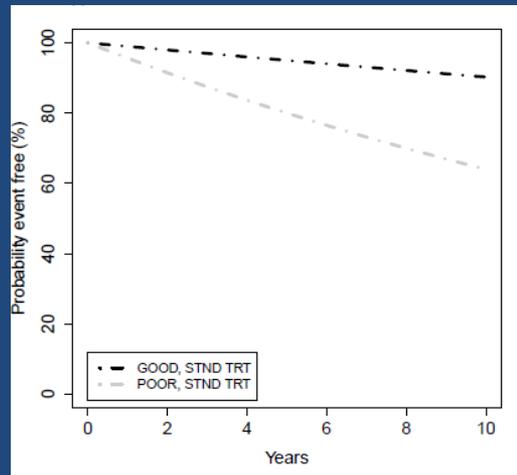
- ❑ Clear intended use with clinical utility
- ❑ Is a prospective trial needed, and if so, what design?
- ❑ Protocol with clear objectives, design, statistical analysis plan, locked down predictor
- ❑ Secure database
- ❑ Responsible individuals named

# Domain 4: Prognostic ability only sometimes translates to clinical utility

Is this prognostic information helpful ?



Good prognosis group may forego additional therapy



## Domain 4: Usually we are more interested in “predictive” ability of a biomarker or omics predictor

- Predictive: Associated with benefit or lack of benefit (potentially even harm) from a particular therapy relative to other available therapy
  - Alternate terms: treatment-selection, treatment-guiding, treatment effect modifier
- Need randomized treatment trial, or at least specimens collected from such a trial

Polley et al, J Natl Cancer Inst 2013;105:1677-1683  
McShane & Polley, Clinical Trials 2013; 10: 653-665

# Domain 4: Study designs to examine tests/biomarkers for guiding therapy

## □ Main types of prospective designs

- Biomarker-Enrichment; Biomarker-Strategy; Biomarker-Stratified; All-comers

Sargent D et al. *J Clin Oncol* 2005;23:2020-2027

Freidlin B et al., *J Natl Cancer Inst* 2010;102:152-160

Clark G & McShane L, *Stat Biopharm Res* 2011;3:549-560

## □ Prospective-retrospective design

- Stored specimens from completed prospective trial
- Clear pre-specified study objectives
- Rigorous statistical design & analysis plans

Simon et al, *J Natl Cancer Inst* 2009;101:1446-1452

## Domain 5: Ethical, legal, and regulatory issues

- ❑ Informed consent discloses investigational use, risks, potential COIs
- ❑ Intellectual property
- ❑ Requirements for tests to be performed in CLIA-certified laboratory
- ❑ Determine if investigational device exemption (IDE) is required from FDA

# Case study: Serum proteomic test to guide use of EGFR-TKI therapy for patients with lung cancer

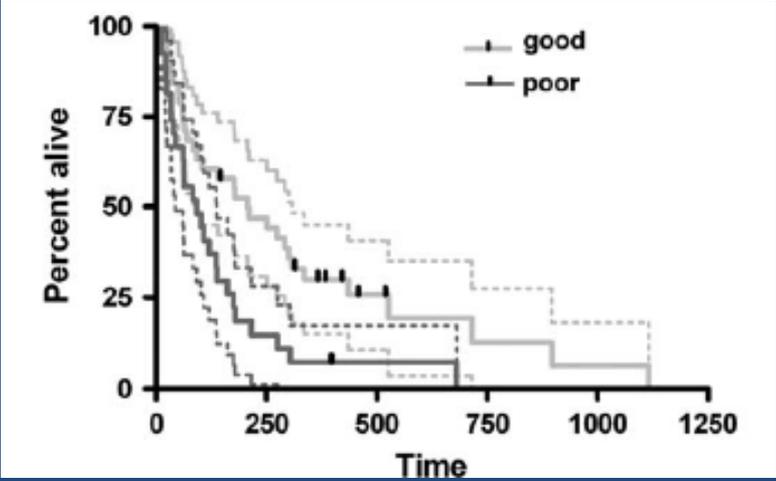
- ❑ Patients with advanced non-small cell lung cancer typically have poor outcome with standard chemotherapies
- ❑ Some new drugs have been designed to be effective against tumors that have alterations in the EGFR gene (EGFR-TKIs)
- ❑ Determination of whether a tumor has an EGFR alteration has traditionally required obtaining a biopsy of the tumor
- ❑ A serum proteomic test, if proven reliable, could avoid the need for tumor biopsy to evaluate likelihood of sensitivity to EGFR-TKIs

# Model development for serum proteomic test

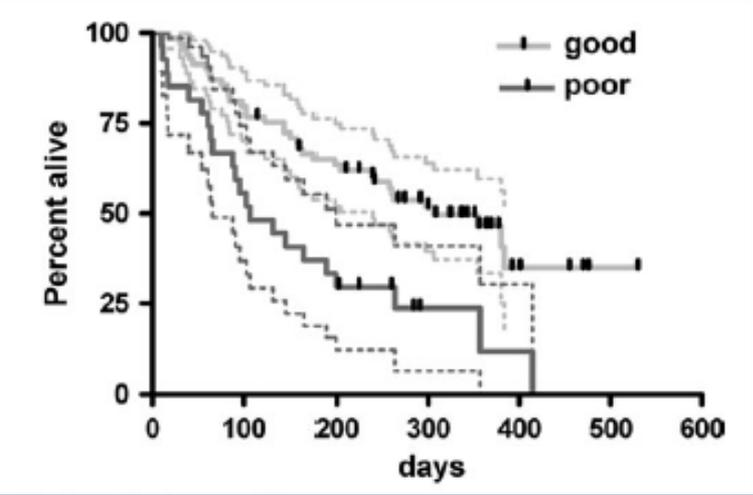
- ❑ Serum collected from NSCLC patients before treatment with gefitinib or erlotinib (EGFR-TKIs)
- ❑ Analysis by MALDI-MS
- ❑ K-nearest neighbor (KNN) algorithm based on 8 distinct m/z features classifies into good or poor outcome
- ❑ Training set: n=139 NSCLC patients total from 3 cohorts who received gefitinib
- ❑ Preliminary validation cohorts:
  - “Italian B”: n=67 sequential patients, late-stage or recurrent NSCLC treated with single-agent gefitinib
  - ECOG 3503: n=96 advanced NSCLC patients treated with first-line erlotinib on single arm Phase II study

# Initial assessment of serum proteomic test

Preliminary results for patients treated with EGFR-TKIs



“Italian B”: n=67 sequential patients, late-stage or recurrent NSCLC treated with single-agent gefitinib  
HR\*=0.50, 95% CI=(0.24,0.78), p=0.0054  
Median OS  
Good: 207 days Poor: 92 days



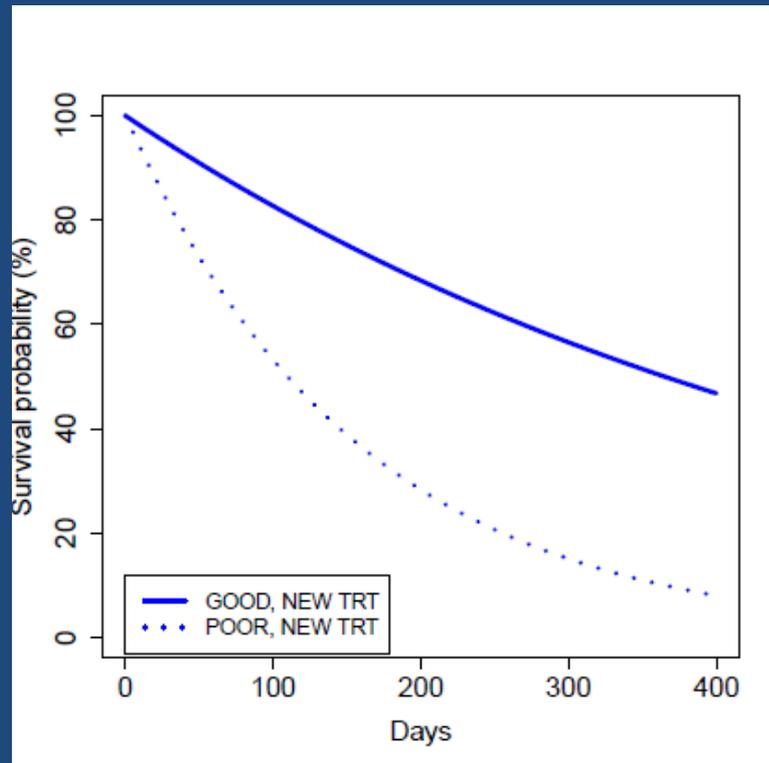
ECOG 3503: n=96 advanced NSCLC patients treated with first-line erlotinib on single arm Phase II study  
HR\*=0.4, 95% CI=(0.24,0.70), p<0.001  
Median OS  
Good: 306 days Poor: 107 days

In addition, proteomic test shown to have good analytical reproducibility across 2 labs

\*HR for Good:Poor

# Serum proteomic test: Predictive or prognostic?

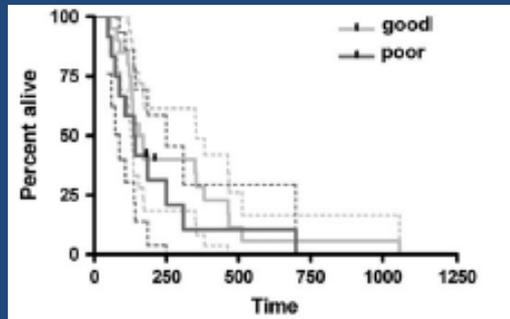
This is what we see for patients who received the EGFR-TKIs



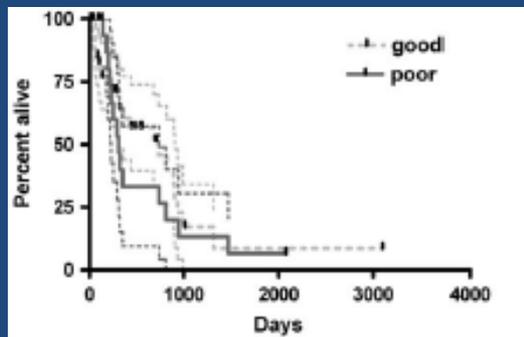
BUT, what does survival look like for patients who receive standard chemotherapy?

# Serum proteomic test: Predictive or prognostic?

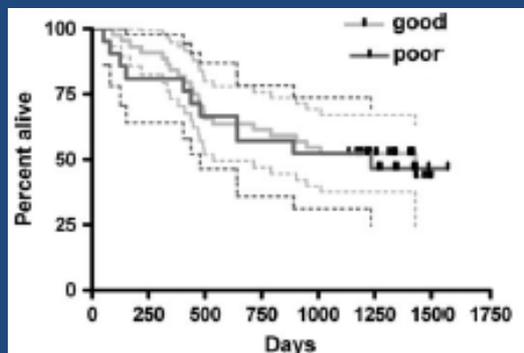
Does test also separate, by outcome, patients who did NOT receive EGFR-TKIs (control cohorts)?



“Italian C”: n=32 patients, stage IIIA-IV NSCLC treated with second-line chemotherapy  
HR\*=0.74, 95% CI=(0.33,1.6), p=0.42  
SAME TREND (HR<1) as in EGFR-TKI treated, but not significant



“VU”: n=61 patients, advanced NSCLC treated with second-line chemotherapy  
HR\*=0.81, 95% CI=(0.4,1.6), p=0.54  
SAME TREND (HR<1) as in EGFR-TKI treated, but not significant



“Polish”: n=65 patients, stage IA-IIB NSCLC treated with second-line chemotherapy  
HR\*=0.90, 95% CI=(0.43,1.89), p=0.79  
SAME TREND (HR<1) as in EGFR-TKI treated, but not significant  
\*HR for Good:Poor

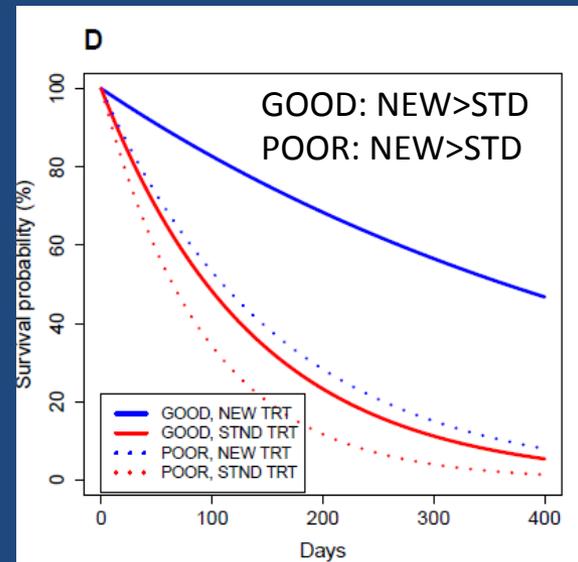
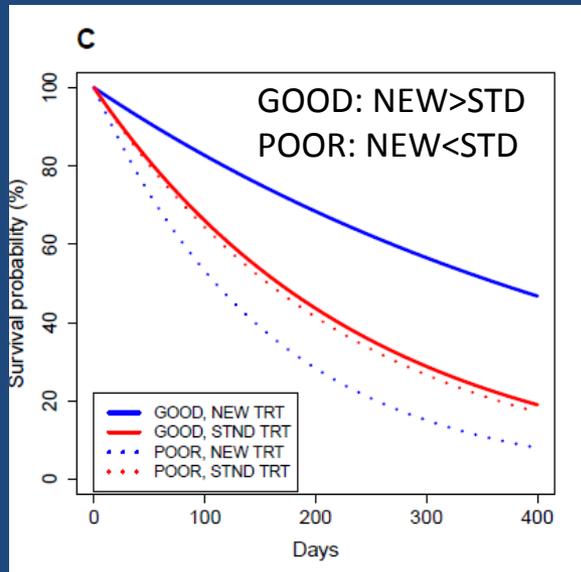
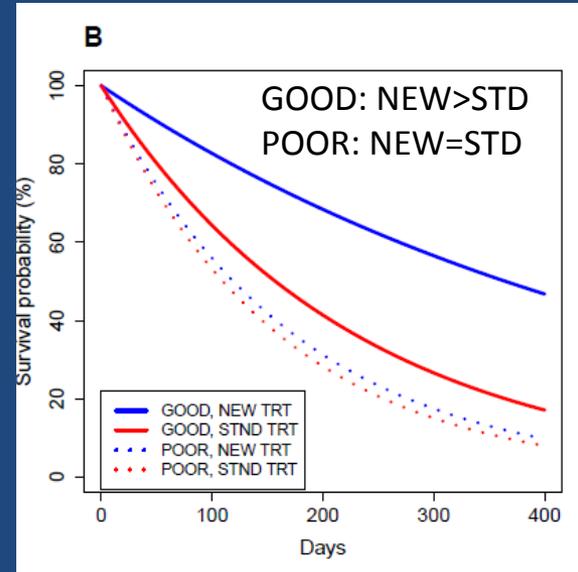
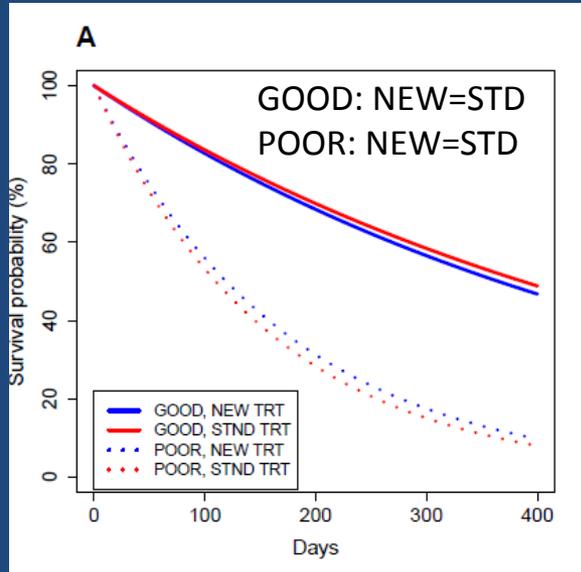
# Serum proteomic test: Need a randomized clinical trial to draw conclusions

- ❑ Therapy not randomized in the retrospective studies used for development
- ❑ Clinical characteristics (e.g., stage) differed across the patient cohorts

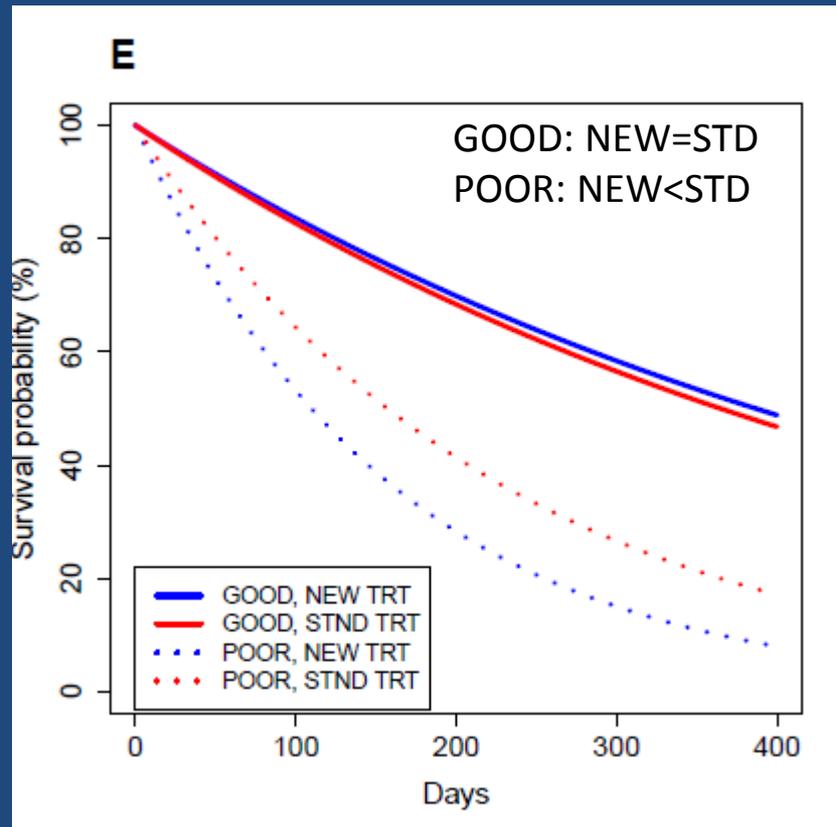
# Randomized phase III trial (PROSE) to evaluate ability of serum proteomic test to predict benefit from EGFR-TKIs

- ❑ Test predictive value of the proteomic test
- ❑ Primary endpoint overall survival (OS)
- ❑ Powered for treatment x proteomic test interaction (biomarker-stratified design)
- ❑ Eligibility
  - Stage IIIB or IV NSCLC
  - $\geq 18$  years old
  - Refractory to one previous platinum-containing regimen
- ❑ Exclusions
  - Previously received an EGFR-TKI
  - Uncontrolled brain metastases
  - Other cardiac, renal, etc. conditions

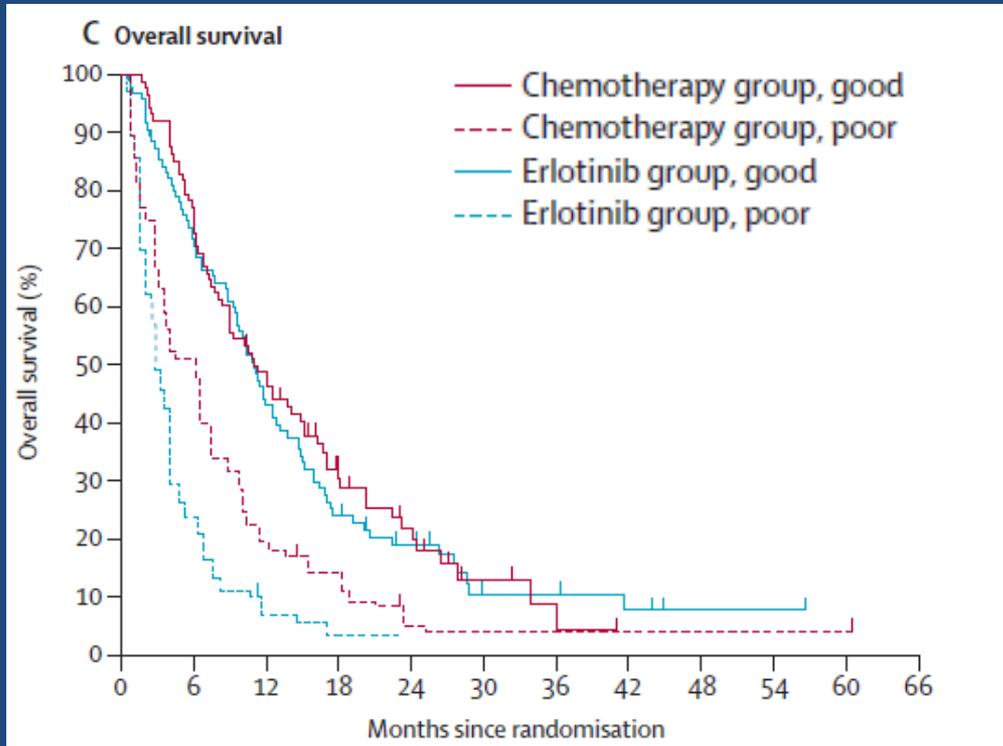
# Serum proteomic test: Some possible clinical trial outcomes



# Serum proteomic test: The real clinical trial outcome was none of the above



# PROSE trial results for overall survival



## Median Overall Survival (mos.)

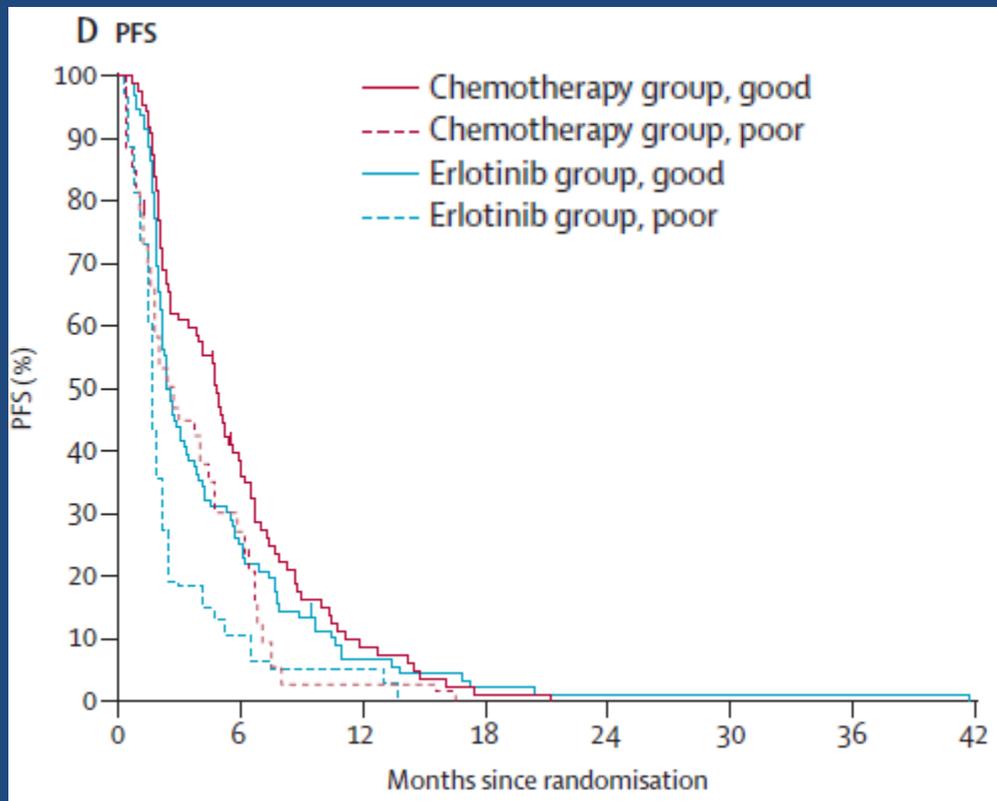
	Test result	
Treatment	Good	Poor
Chemo	10.9	6.4
Erlotinib	11.0	3.0
Hazard ratio*	1.06	1.72
(95% CI)	(0.77-1.46)	(1.08-2.74)

Interaction  $p=0.017$

\*HR for Erlotinib:Chemo

Not even a trend for better outcome with erlotinib in the “good” group.

# PROSE trial results for progression-free survival



## Median Progression-Free Survival (mos.)

<u>Treatment</u>	<u>Test result</u>	
	Good	Poor
Chemo	4.8	2.8
Erlotinib	2.5	1.7
Hazard ratio*	1.26	1.51
(95% CI)	(0.94-1.96)	(0.96-2.38)

Interaction  $p=0.445$

\*HR for Erlotinib:Chemo

Trend for longer PFS with chemotherapy in the “good” group.

# PROSE trial results

Conclusion drawn by authors:

“Serum protein test status is predictive of differential benefit in overall survival for erlotinib versus chemotherapy in the second-line setting. Patients classified as likely to have a poor outcome have better outcomes on chemotherapy than on erlotinib.”

(Gregorc et al, *Lancet Oncol* 2014;15:713-721)

Is this test clinically useful?

# Summary remarks

- ❑ Scientific teams that develop omics tests should include individuals with statistical expertise
- ❑ Familiarize yourself with checklists and reporting guidelines BEFORE you start your study
- ❑ Statisticians have a responsibility to engage in the scientific process and not naively churn out statistical analyses

THANK YOU!  
lm5h@nih.gov